Synthesis of alkylated sugar amino acids: conformationally restricted L-Xaa-L-Ser/Thr mimics†

Martijn D. P. Risseeuw,[a] Jaroslaw Mazurek,[b] Arjan van Langenvelde,[b] Gijsbert A. van der Marel,[a] Herman S. Overkleeft[a] and Mark Overhand*[a]

Two synthetic strategies for the generation of δ-substituted pyranoid sugar amino acids (SAAs) are evaluated. The first employs chiral nonracemic *tert*-butane sulfinamides as key reagents. Regardless of the stereochemistry of the applied sulfinamide, the product formed has a stereochemistry resembling that of a D amino acid at C7. Direct Grignard reaction on formyl-tetra-*O*-benzyl-β-D-*C*-glucopyranoside in the second strategy and subsequent Mitsunobu inversion, yields the L,L-dipeptide isosters.

Introduction

As part of our ongoing research on artificial peptide-like materials, we recently reported the synthesis of glucopyranose-based sugar amino acids (SAAs) **1** (Fig. 1).[1] In dipeptide isosteres **1**, the stereocentre at C2 has the *S*-configuration, thereby resembling the α-carbon in L-serine or L-threonine.[2] The secondary amine at the N-terminus (C7) in **1** resembles the side-chain at the α-carbon of amino acids other than glycine. By tuning the nature of the R[1] group, functionalities corresponding to specific amino acid side-chains can be incorporated into the SAAs.[3] This feature distinguishes compounds **1** from SAAs reported in the literature,[4] of which the large majority are primary amines.[5] As a whole, SAAs **1** can be viewed as conformationally constrained H-Xaa-Ser/Thr-OH mimics. The configuration of C7 corresponds to that of the α-carbons in D-amino acids, rather than the proteinogenic L-amino acids. In contrast to stereochemical control over the C-terminal portion, which originates from selection of the carbohydrate template, controlling the configuration of the newly introduced stereocentre at C7 stems from asymmetric organic synthesis. The work presented here concerns adaptation of the synthetic strategy we applied to prepare SAAs **1** to provide C7-*S* configured SAA building blocks **2**.

[a]Leiden Institute of Chemistry, Department of Bioorganic Synthesis, PO box 9502, 2300 RA, Leiden, The Netherlands. E-mail: overhand@chem.leidenuniv.nl
[b]Avantium Technologies, Zekeringstraat 29, 1014BV, Amsterdam, The Netherlands

† Electronic supplementary information (ESI) available: Experimental details. See DOI: 10.1039/b705750d

Protected SAAs **1** were previously prepared using the stereoselective alkylation of *R-tert*-butanesulfinimide **4**, which was obtained by the condensation of formyl tetra-*O*-benzyl-β-D-*C*-glucopyranoside **3**[6] with *R-tert*-butanesulfinyl amide[7] (Scheme 1). Alkylation of compound **4**, subsequent acid-mediated hydrolysis of the *R-tert*-butanesulfonyl group and instalment of the Fmoc protective group gave compound **7**, which could be transformed into carboxylate **8** by selective acidolysis of the primary benzyl ether, ester hydrolysis and oxidation. The alkylation of sulfinimide **4** proceeds in good diastereoselectivity. For instance, reaction of **4** with 3 equivalents of MeMgBr in methylene chloride at −78 °C gave *R*-methyl adduct **5** in 20-fold excess over the other diastereoisomer **6**. Similar results were obtained by using toluene as a solvent. Performing the alkylation in THF resulted in a drop in diastereoselectivity (**5**–**6** = 13 : 1).

Either the *R-tert*-butanesulfinyl chiral auxiliary or the chiral carbohydrate template, or a combination of both may be responsible for the observed diastereoselectivity. Would the first be true, then L,L-dipeptide isostere SAAs **2** would be directly accessible by following the same synthetic scheme, but employing *S-tert*-butanesulfinimide as the chiral auxiliary. In order to investigate this possibility, we prepared *S-tert*-butanesulfinimide **9**, the diastereoisomer of **4** with respect to the chirality at the sulfur atom. Treatment of *S-tert*-butanesulfinimide **9** with 3 equivalents of MeMgBr (CH$_2$Cl$_2$, −78 °C) gave a diastereoisomeric ratio for **10**–**11** of 13 : 1, as monitored by inverse gated ^{13}C NMR measurements[8] on the crude Grignard products (Scheme 1). The absolute stereochemistry of **10** was unambiguously established by acidic removal of the *S-tert*-butanesulfonyl group, giving, after Fmoc-protection, a compound that in all spectroscopical
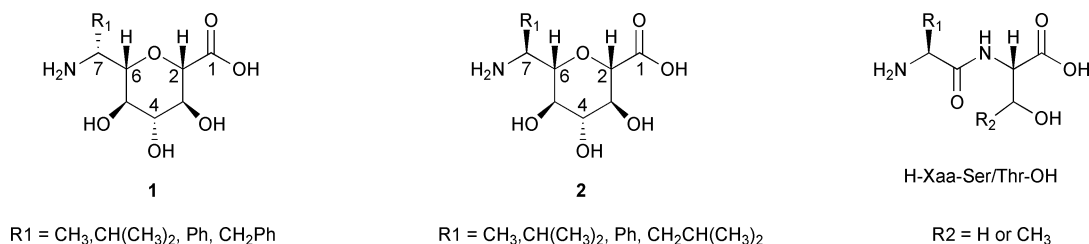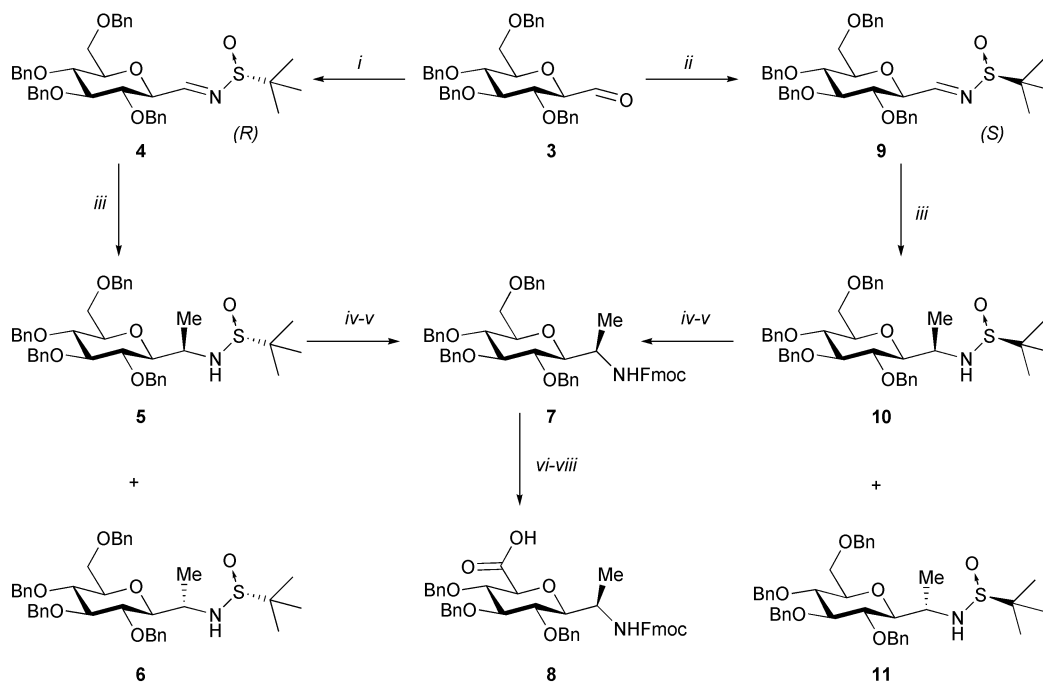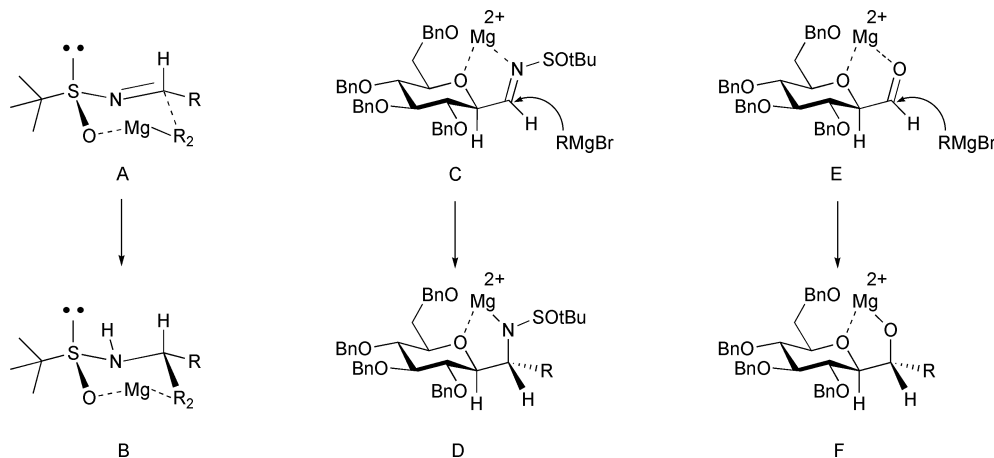

Fig. 1 Alkylated SAAs as Xaa-Ser/Thr mimics.

1   R1 = CH$_3$, CH(CH$_3$)$_2$, Ph, CH$_2$Ph

2   R1 = CH$_3$, CH(CH$_3$)$_2$, Ph, CH$_2$CH(CH$_3$)$_2$

H-Xaa-Ser/Thr-OH

R2 = H or CH$_3$

Scheme 1 *Reagents and conditions:* (i) *R-tert*-butanesulfinamide, Ti(OiPr)₄, CH₂Cl₂, 70%, (ii) *S-tert*-butanesulfinamide, Ti(OiPr)₄, CH₂Cl₂, 72%, (iii) MeMgBr, CH₂Cl₂, −78 °C, (iv) HCl, MeOH, (v) FmocOSu, DIPEA, dioxane, CH₂Cl₂, (from **4**: 71%, from **9**: 75%, three steps), (vi) ZnCl₂, HOAc, Ac₂O, (vii) HCl, MeOH, (viii) TEMPO, BAIB, CH₂Cl₂, H₂O (64%, two steps).

aspects was identical to the previously synthesized **7** (R¹ = CH₃).[1] From this it follows that the minor product **11** is the *S*-methyl diastereomer of **10** with respect to the newly formed stereocentre. Apparently, *re*-side addition is favored irrespective of the nature of the chiral auxiliary on the imine nitrogen. Changing the solvent system from CH₂Cl₂ to THF resulted in a slightly more favored *si*-side addition, and **10** and **11** were formed in equal amounts. This result is the best we obtained in favor of the desired diastereoisomer **11** and we conclude that at least in this system chiral sulfinylimides are not useful intermediates in the construction of ʟ,ʟ-dipeptide isosteres.

At the moment we do not have a satisfying model that explains the different product ratios we observe, but it seems likely that the chelation model proposed by Ellman and co-workers in their explanation[7f] of chirality transfer (**A** to **B**, Fig. 2) is counterbalanced by competing chelation of magnesium ions to hetero-atoms on the carbohydrate template. This chelation (for instance **C**, leading to **D**) may occur irrespective of the configuration on the sulfur atom. Whether this reasoning is valid or not, it does present an obvious strategy towards the desired ʟ,ʟ-dipeptide isosteres. When one assumes that a formyl-*C*-glycoside chelates just as the sulfinyl imides do with magnesium and that

Fig. 2 Transition states for Grignard reactions (**A** → **B**: sulfinimines in general, **C** → **D**: glycosyl sulfinimide through intramolecular chelation, **E** → **F**: glycosyl aldehyde through intramolecular chelation).

Scheme 2 *Reagents and conditions:* (i) PhMgBr, THF, −78 °C (66%), (ii) RMgBr or RMgCl, THF, 0 °C (**a** 66%; **b** 51%; **c** 36%; **d** 48%), (iii) HN$_3$, DEAD, PPh$_3$, toluene (**a** 78%; **b** 83%; **c** 48%; **d** 92%), (iv) ZnCl$_2$, HOAc, Ac$_2$O, (v) NaOMe, MeOH (**a** 73%; **b** 78%; **c** 68%; **d** 81%, two steps), (vi) TEMPO, PhI(OAc)$_2$, CH$_2$Cl$_2$, H$_2$O (**a** 89%; **b** 91%; **c** 76%; **d** 92%), (vii) first Me$_3$P, THF, H$_2$O, then FmocCl, CH$_2$Cl$_2$–dioxane, DIPEA (73%), (viii) H$_2$, Lindlars catalyst, Boc$_2$O, MeOH (85%), (ix) Pd/C, H$_2$, MeOH, (x) TFA (97%, two steps).

addition occurs with the same *re*-selectivity (**E** to **F**), then the target compounds are within reach by introduction of a nitrogen substituent by replacement of the resulting alcohol function with concomitant reversal of configuration. This reasoning proved to be valid, as is outlined in Scheme 2.

Grignard addition of 3 equivalents of either PhMgBr, MeMgBr, iPrMgCl or iBuMgBr to aldehyde **3** proceeded in good diastereoselectivity to give **12a–d**, respectively, as the single isolated diastereomers in moderate to good yields (Scheme 2). The absolute configuration of the newly formed stereocentre in alcohol **12b** could be assigned as *R* since its analytical data were in excellent agreement with published values.[9] The crystals obtained from recrystallization of compound **12d** proved to be suitable for an X-ray structural determination to show the anticipated (*R*)-configuration at the newly created stereocentre.[10]

Mitsunobu displacement of the secondary alcohols **12a–d** with azide (HN$_3$, PPh$_3$, DEAD, toluene)[11] gave, with inversion of configuration, azides **13a–d**. Of these, phenylglycine analogue **13a** was transformed by a three step sequence (reduction of the azide with concomitant Boc protection yielding derivative **17** followed by hydrogenolytic cleavage of the benzylethers and TFA treatment) into known glucosylamine derivative **18**.[12] All analytical data on **18** are in agreement with those reported in the literature for the

same compound hereby validating the assignment of the newly formed stereocentre in azide **13a**. The structural integrity of our compounds was further established by transformation of azide **13b** into the corresponding Fmoc-protected amide **16** (first Staudinger reduction, then treatment with fluoronylmethyloxycarbonyl chloride and DIPEA), which gave a compound with the same mass but with otherwise distinct spectroscopical properties from *R*-configured *C*-glycoside **7**. Selective acidolysis of the primary benzyl ether and subsequent oxidation of the resulting primary hydroxyls gave, after ester hydrolysis, the L,L-dipeptide isosteres **15a–d**.

In conclusion, we have now at our disposal two related synthesis strategies that enable us to prepare both D,L and L,L glucopyranose derived Xaa–Ser(Thr) dipeptide isosteres. We are currently investigating their structural properties in linear and cyclic homo- and hetero-oligomers, and are pursuing adaptation of the synthesis strategy to also incorporate other functionalised amino acid side-chains.

Experimental

Full experimental procedures and physical data of the compounds described in this paper can be found in the ESI.†

Crystallography

Compound **12d** was crystallized from mixture of diethyl ether and light petroleum at room temperature as parallelepiped blocks.‡ A crystal of approximately $0.6 \times 0.35 \times 0.15$ mm was cut from a larger one and analyzed on the Kappa CCD at 293 K using MoKα_1 radiation. The full sphere was collected up to $\theta = 27.5°$. The data collection and processing were done using the Scalepack software.[13,14] The structure was solved by the direct method and refined using the SHELX97 package.[15] All of the non-H atoms were refined anisotropically while all H atoms were found on the Fourier Difference map and refined isotropically. The absolute configuration of the molecule was deducted knowing the absolute configurations of the four stereocentres at C2, C3, C4 and C5 as occurring in D-glucose, and the Flack parameters.[16]

Acknowledgements

References

1 M. Raunkjaer, M. F. El Oualid, G. A. von der Marel, H. S. Overkleeft and M. Overhand, *Org. Lett.*, 2004, **6**, 3167–3170.

2 E. Graf von Roedern and H. Kessler, *Angew. Chem., Int. Ed. Engl.*, 1994, **33**, 687–689.

3 For related compounds see: (*a*) T. K. Chakraborty and G. Sudhakar, *Tetrahedron Lett.*, 2005, **46**, 4287–4290; (*b*) T. K. Chakraborty and G. Sudhakar, *Tetrahedron: Asymmetry*, 2005, **16**, 7–9; (*c*) S. Schröder, A. K. Schrey, A. Knoll, P. Reiss, B. Ziemer and U. Koert, *Eur. J. Org. Chem.*, 2006, 2766–2776; (*d*) J. E. Campbell, E. E. Englund and S. D. Burke, *Org. Lett.*, 2002, **4**, 2273–2275.

4 For general reviews on SAAs see: (*a*) S. A. W. Gruner, E. Lorcardi, E. Lohof and H. Kessler, *Chem. Rev.*, 2002, **102**, 491–514; (*b*) T. K. Chakraborty, S. Ghosh and S. Jayaprakash, *Curr. Med. Chem.*, 2002, **9**, 421–435; (*c*) F. Schweizer, *Angew. Chem., Int. Ed.*, 2002, **41**, 230–253; (*d*) T. K. Chakraborty, P. Srinivasu, S. Tapadar and B. K. Mohan, *J. Chem. Sci.*, 2004, **116**, 187–207; (*e*) T. K. Chakraborty, P. Srinivasu, S. Tapadar and B. K. Mohan, *Glycoconjugate J.*, 2005, **22**, 83–93.

5 For some examples of sugar amino acids possessing a primary amine functionality see: (*a*) S. F. Jenkinson, T. Harris and G. W. J. Fleet, *Tetrahedron: Asymmetry*, 2004, **15**, 2667–2679; (*b*) T. K. Chakraborty, S. Ghosh, S. Jayaprakash, J. A. R. P. Sharma, V. Ravikanth, P. V. Diwan, R. Nagaraj and A. C. Kunwar, *J. Org. Chem.*, 2000, **65**, 6441–6457; (*c*) H. S. Overkleeft, S. H. L. Verhelst, E. Pieterman, N. J. Meeuwenoord, M. Overhand, L. H. Cohen, G. A. von der Marel and J. H. van Boom, *Tetrahedron Lett.*, 1999, **40**, 4103–4106; (*d*) F. Perl, L. Cipolla, B. La Feria and F. Nicotra, *Chem. Commun.*, 2000, 2303–2304.

6 (*a*) W. R. Kobertz, C. R. Bertozzi and M. D. Bednarski, *Tetrahedron Lett.*, 1992, **33**, 737–740; (*b*) A. Dondoni and M.-C. Scherrmann, *J. Org. Chem.*, 1994, **59**, 6404–6412; (*c*) M. E. Sánchez, V. Michelet, I. Besnier and J. P. Genêt, *Synlett*, 1994, 705–708; (*d*) F. Labeguere, J. P. Lavergne and J. Martinez, *Tetrahedron Lett.*, 2002, **43**, 7271–7272; (*e*) A. Dondoni and A. Marra, *Tetrahedron Lett.*, 2003, **44**, 13–16.

7 For the application of enantiopure *tert*-butanesulfinamides in the generation of chiral secondary amines see: (*a*) G. C. Liu, D. A. Cogan and J. A. Ellman, *J. Am. Chem. Soc.*, 1997, **119**, 9913–9914; (*b*) A. Lee and J. A. Ellman, *Org. Lett.*, 2001, **3**, 3707–3709; (*c*) T. P. Tang and J. A. Ellman, *J. Org. Chem.*, 2002, **67**, 7819–7832; (*d*) T. P. Tang, S. K. Volkman and J. A. Ellman, *J. Org. Chem.*, 2001, **66**, 8772–8778; (*e*) D. J. Weix and J. A. Ellman, *Org. Lett.*, 2003, **5**, 1317–1320; (*f*) J. A. Ellman,

Pure Appl. Chem., 2003, **75**, 39–46; (*g*) D. Morton and R. A. Stockman, *Tetrahedron*, 2006, **62**, 8869–8905.

8 Inverse gated ^{13}C NMR is a technique that allows quantitation of carbon atoms by using longer relaxation times and removal of any NOE enhancements. For more information see: T. D. W. Claridge, in *High-Resolution NMR Techniques in Organic Chemistry*, Pergamon, Oxford, 1999, ch. 4, pp. 111–146.

9 C. F. Brewer, E. J. Hehre, J. Lehmann and W. Weiser, *Liebigs Ann. Chem.*, 1984, 1078–1087.

10 Ortep representation of the X-ray structure of compound **12d**. Thermal ellipsoids are set for 35%..
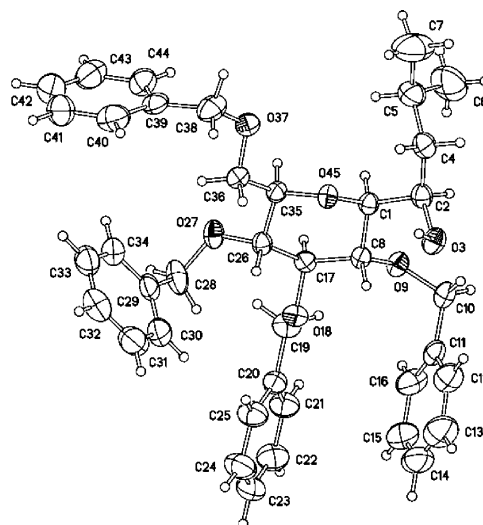
Empirical formula	$C_{39}H_{46}O_6$
Formula weight	610.76
T[K]	293(2)
λ[Å]	0.71073
Crystal system	Orthorhombic
Space group	P $2_1 2_1 2_1$
Unit cell dimensions	
a [Å]	6.0520(2)
b [Å]	23.7560(5)
c [Å]	23.9440(5)
V [Å^3]	3442.5(2)
Z	4
D_m [g/cm^3]	1.178
Absorption coefficient mm^{-1}	0.078
F(000)	1312
Crystal size [mm^3]	0.6 x 0.3 x 0.15
θ range	3 → 27.5
Reflections collected	17437
Independent reflections	7673 [R_{int} = 0.0411]
Data / restraints / parameters	7673 / 0 / 566
S	1.025
R [I>2σ(I)]	R1 = 0.0465, wR2 = 0.0858
R indices (all data)	R1 = 0.0747, wR2 = 0.0965
Absolute structure parameter	-0.4(7)

11 T. Mukaiyama, I. Shiina, H. Iwadare, M. Saitoh, T. Nishimura, N. Ohkawa, H. Sakoh, K. Nishimura, Y. Tani, M. Hasegawa, K. Yamada and K. Saitoh, *Chem.–Eur. J.*, 1999, **5**, 121–161.

12 R. R. Schmidt and H. Dietrich, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 1328–1329.

13 S. Mackay, C. J. Gilmore, C. Edwards, N. Stewart and K. Shankland, *maXus*, Bruker Nonius, The Netherlands, MacScience, Japan & The University of Glasgow, 1999.

14 Z. Otwinowski and W. Minor, *Methods Enzymol.*, 1997, **276**, 307–326.

15 (*a*) G. M. Sheldrick, SHELXS-97: Program for Crystal Structure Solution, University of Göttingen, Germany, 1997; (*b*) G. M. Sheldrick, SHELXL97: Program for the Refinement of Crystal Structures, University of Göttingen, Germany, 1997.

16 H. D. Flack, *Acta Crystallogr., Sect. A*, 1983, **39**, 876–881.

2314 | *Org. Biomol. Chem.*, 2007, **5**, 2311–2314

This journal is © The Royal Society of Chemistry 2007